EVALUATION OF THE EFFICIENCY OF CERTAIN BIO-AGENT COMPOUNDS COMPARED WITH THE NEMATICIDE, CADUSAFOS AGAINST R00T - KNOT NEMATODE, *Meloidogyne incognita*

Mahgoob¹, Ahmed E.A; Sayed M. Dahroug^{1;} Ahmed A. A. Deabes² and Aida M. El-Khouly²

Plant Protection Department, Faculty of Agriculture, Ain Shams University.
 Fungicides, Bactericides and Nematicides Dept. - Central Agricultural Pesticides Lab., ARC.

ABSTRACT: The present study was conducted to evaluate the effectiveness of some bio-agent compounds, e. g., Sincocin, Nematoxin, Bio-Nematon, Nembecidin, and Bio-Nema comparing with the nematicide, cadusafos (10% G) against the root-knot nematode, M. incognita under laboratory and green house conditions. After one day exposure period, cadusafos and Nembecidin caused the lowest percentage of hatchability at double rate of application, while the highest percentage was at quarter rate of application of them, respectively. After 7 days, Nematoxin and Bio-Nematon were the most efficiency at double rate of application of them, respectively. Whereas the least efficiency was at quarter and double rate of application of Sincocin, respectively and quarter rate of application of Bionematon. In respect of the mortality of second stage juveniles, the highest mortality percentage after one day exposure period was at double rate of application of Nembecidin followed by cadusafos, while lowest one was at quarter rate of application of them. Similar trend were observed after 7 days of exposure period of the rest of the tested bio-agent compounds. However, Bio-Nema and Nematoxin were the most efficiency while the lowest ones were Bio-Nematon, Bio-Nema and Sincocin,. Regarding effect of application time and rate on the effectiveness of the tested bio-agent compounds against M. incognita, Nematoxin was the most efficiency where caused the highest percentage of reduction for the numbers of galls, J2s in soil and total number of eggs while treatment with Sincocin resulted the lowest one for the same parameters, this data when application time was after nematode infection. Treatment with Bio-Nematon resulted the highest percentage of reduction for the number of galls, J2s in soil and total number of eggs whereas treatment with Sincocin sighted the lowest ones for the same parameters, this when application time was before nematode infection. Treatment with Bio-Nema when applied before nematode infection increased the length of shoot, root and the fresh weight of shoot and root in addition of total number of leaves, whereas decreased the same parameters as a percentage when applied after nematode infection, this comparing with the control (inoculated untreated).

Keywords: Plant parasitic nematodes, bio-agent compounds, nematicides.

1. INTRODUCTION

The plant parasitic nematodes caused great economic losses to agricultural crops worldwide (**Sasser, 1979**) and are considered an economically important group of soil borne pathogens. Since nematodes are of great economic importance, much attention has been paid to their control. Root-knot nematodes *Meloidogyne incognita* (Kofoid & White) are serious pathogen of most crops causing quality defects that result in reduced crop value or crop rejection particularly in the warm climate. Root-knot nematodes cause annual losses of about 100 billion USD worldwide (**Brand** *et al.*, **2010**). Among various control measures, the use of nematicides has been found to be highly effective and reliable in controlling a wide range of

nematodes. On the other hand, chemical control is expensive and their residues persist in the environment for a longer period. Also, affects the beneficial soil microorganisms. Non fumigant nematicides costs are approximately 120- 220 \$ / acre. All registered non fumigant nematicides are carbamates or organophosphates and 1, 3 - dichloropropene are second carcinogenic grade. All the three compounds are at the top of the "hit list" developed to implement the Food Quality Protection Act. Finally, development of natural resistance to nematicides by nematodes and the tendency to withdraw chemical pesticides/nematicides from the market led to the search for new methods of control. Hence, several biological control agents processing nematicidal properties were investigated as possible measures for controlling the nematode disease in various crops. Therefore, there is still need for alternatives and environmentally friendly compounds for effective nematode control. Certain safe procedures for nematode control have been developed based on bio-control agents and organic amendments, control plant parasitic nematode by reducing egg production of root - knot nematode which infects roots of several hosts (Jatala et al., 1985; Jatala 1986; Prakob et al., 2007 and Shahnaz et al., 2008). Research on the possible mechanisms involved in biological control by bio-agents has led to several alternative explanations for successful bio control. Based on this background, greenhouse experiments aimed at substituting nematicides in controlling root-knot disease of okra plants induced by Meloidogyne spp. using of certain bio-agent compounds. The present work was carried out to evaluate five bio agent compounds namely, Bio--Nema, Bio-Nematon, Sincocin, Nematoxin and Nembecidin to testify their efficacy on eggs and larvae (J2) for Meloidogyne spp. in vitro as well as their effect on the pathogen root- knot nematode, Meloidogyne incognita and the host (okra plants) under greenhouse conditions to avoid the hazards of chemicals.

2.1. The plant parasitic nematodes.

2.1.1. Root-knot nematode culture:

Culture of *Meloidogyne incognita* was maintained on eggplant plants (*Solanum melongena* cv. Black Beauty) growing in 20 cm diameter clay pots filled with steam sterilized soil comprised of 1 clay: 2 sand (v /v). Infected plants were placed in a greenhouse at 25 ± 5 °C and watered regularly. Two months later, infected roots were cut down into pieces and used as sources of inoculation for other healthy eggplant seedlings. By repeating this procedure, enough quantities of *M. incognita* inocula were obtained for other experiments. Mature egg- masses obtained from the source cultures were put in sterilized water in Petri-dishes for hatching of second stage juveniles (J₂s). The emerged J₂s were collected and refrigerated at 9^oC for the experimental use.

2.1.2. Hatchability of eggs and mortality of J₂s, *M. incognita* treatments:

For testing the nematicidal efficiency of tested bioagent compounds, five rates of each bio-agent compound were prepared: 1/4, 1/2, 1, 2 and 4 rate of application. A volume of 10 ml from each rate was transferred to Petridishes. About1000 eggs or 300 newly hatched second juvenile stage larvae (J2s) of *M. incognita* were transferred to each Petri- dish. Three replicates were used per each rate and free distilled water served as controls. In case of the nematicide, cadusafos and a bio-agent compound, Nembecidin treatments, larval mortality count and egg hatchability were determined after 1 day, while the other compounds were obtained after 7 days, respectively. Percentages of mortality were corrected according to **Abbott (1925).**

Abbott's formula (% mortality) =

Dead in treatment – Dead in control

100 – Dead in control

2. MATERIALS AND METHODS

2.1.3. Determination of nematode parameters:

X 100

Soil of each pot were soaked in a bucket half-full of tap water for 15 min. Nematode extraction was conducted by sieving through 200 and 400 mesh sieves (**Goodey**, **1957**). Nematodes in the suspension were obtained from the 400 mesh and extracted for 72 hours by the modified Baerman trays technique (**Goody**, **1963**). The final volume of extracted suspension was adjusted to about 50 ml. The nematodes were counted using Hawksley counting slide and light microscope. The number of galls or/and egg masses per each root system were counted using dissecting microscope. Regarding estimation of the average number of eggs per an eggmass, ten egg masses were selected randomly and put in a 1% NaOCI solution for 3 min. with shaking, then the suspension of eggs was sieved through 200 and 500 mesh sieves with gentle tap water to remove the debris by the first sieve and collected the released eggs by the second one (**Hussey and Barker**, **1973**), average number of eggs per an egg mass was calculated.

2.2. Determination of plant parameters:-

Shoot & root length (cm), shoot & root fresh weights (gm.) and the number of leaves /plant were counted.

2.3. Bio-agent compounds and the nematicide used:

Five tested bio-agent compounds which not registed in Egypt as nematicides and a registed nematicide, cadusafos were illustrated in Table (1).

Commercial name	Essential components / Concentration	The rate of application
Bio-Nema	<i>Bacillus penetrans</i> (32x10 ³ spores /g) Liquid	10 L / feddan
Bio-Nematon	Paecilomyces Lilacinus ($1x10^9$ CFU's/ml) - Liquid	2 L / feddan
Nematoxin	Plant extracts 25% - Liquid	1.5 L / feddan
Nembecidin.	Azadirachtin 0.03 % – EC.	1 L / feddan
Sincocin	Plant extracts of <i>Rhizopgora mangle</i> , <i>Rhus aromatic</i> (sumac), <i>Quercus falcate</i> , <i>Opunita lindheimeria (Prickly Peasrcactus)</i> and (Magngrove) – Liquid.	0.4 L / feddan
Rugby (Common name: cadusafos)	S, S-di-sec-butyl O-ethyl phosphorodithioate (10 %) - Granules.	20kg / feddan

Table (1): The tested bio- agent compounds and the nematicide, cadusafos.

2.4. Green house experiments:

To study the effect of time and the rate of applications on the efficiency of the tested compounds against rootknot nematode, *M. incognita* on okra plants, the following conducted experiments. Five seeds of okra plants (*Hibiscus esculentus*, cv. Balady) were sowed in each pot then seedlings were thinned after ten days to one/pot. Four replicates were used for each treatment. Each pot was inoculated with about 2000 newly hatched second procedures were carried out using clay pots (15 cm diameter). Soil was autoclaved before used in the experiment. Autoclaved soil was used as 1 kg/pot in all stage juveniles (J2s) of *M. incognita* by pipetting the inoculum suspension into 5 cm deep holes around the rhizosphere. Immediately after inoculation the holes and roots were covered with soil and watered. The proper amount of tested bio-agent compound was added to each

pot according to the experimental aim. For testing the effect of application time and rate on the effectiveness of the bio-agent compounds and cadusafos (10% g) against *M. incognita*, The tested bio-agent compounds were used at three rates, R (Recommended rate); 1/2 R and 2 R. These tested compounds were applied one week after or before nematode inoculation. The plants were plucked after 45 days from inoculation. Numbers of galls, egg masses per root system and number of second stage larvae in the soils were counted. Average number of eggs per an egg mass was calculated. Also the plant growth parameters were determined; shoot & root length (cm), shoot & root fresh weights (gm.) and the number of leaves /plant were counted.

2.5. Statistical Analysis:

Obtained results were analyzed using proc. ANOVA in SAS Mean Separation which conducted using Duncan's multiple range tests in the same program.

3. RESULT AND DISCUSSION

3.1. Effectiveness of the tested bio-agent compounds on the hatchability of eggs and mortality of J₂s, *M. incognita*.

Data given in Table (2) revealed that, there were a highly difference in the eggs hatchability as affected by the tested bio-agent compounds, Nembecidin and the nematicide, cadusafos. After one day exposure period, the lowest percentages of hatchability were 21.7% and 19.1for cadusafos, and (4 rate of application) of Nembecidin respectively, while, the highest percentages were 43.8 and 40.1 % at (¼ rate of application) of each cadusafos and Nembecidin, respectively. 7 days after application, the lowest percentages were 20.9 and 37.9% at (4 rate of application) of Nematoxin and Bio-nema, respectively. Whereas, the highest percentages resulted in decreasing percentages of eggs hatchability. Data in Table (3) indicated that, the highest mortality percentage after one day exposure period were 89 and 83% at (4 rate

45

of application) of each Nembecidin and cadusafos, respectively. While the lowest ones were 20 and 25% at (1/4 rate of application) of each cadusafos and Nembecidin, respectively. Similar trend was observed after 7 days of exposure period of the rest of the tested bio-agent compounds. However, the highest mortality percentages were 96 and 53% at (4 rate of application) of Nematoxin and Bio-Nema, respectively, while the lowest ones at (1/4)rate of application) were 0.0, 9 and 9% of Bio-Nematon, bio-Nema and Sincocin, respectively. Increasing rates resulted in increasing mortality percentage. It could be concluded that the tested bio-agent compounds and cadusafos achieved double effect on the egg hatchability and mortality of J2s of M. incognita. The obtained results are supported in part by the findings of Khanna and Kumar (2006), which tested or evaluated some formulations such as Neem Jeevan, Neemark, Neem Gold, Achook and Kranti some leaf extracts against M. incognita, in terms of their effect on egg hatchability and juvenile mortality. They found that, significantly reduction in egg hatching and increasing in the mortality percentage of J2s of M. incognita at all the tested levels, in comparison with control. Also the nematicidal activity increased with the increase of the concentration and the exposure period of the nematode. The effect of Nembecidin may be due to its components such as, azadirachtin and other limonoids like meliantriol, salanin, nimbin and the most of other terpinoids in the ratio as it occurs in neem in nature. In respect of Bio-Nematon, there were multitude investigations interpreted the actions of P. lilacinus on plant parasitic nematodes as follow, Jatala et al. (1985) mentioned that, P. lilacinus caused substantial egg deformation in M. incognita, these deformed eggs never matured or hatched and killing juveniles and females of *M. incognita*. In the laboratory test this fungus infects eggs of *M. incognita* and destroys the embryos within 5 days because of simple penetration of the egg cuticle by individual hypha aided by mechanical and/or enzymatic activities (Jatala 1986). Also, P. lilacinus suppressed root- knot infections which

resulted in fewer galls developing in the root system (**Prakob** *et al.*, 2007). The serine protease produced by *P*. *lilacinus* might play a role in penetration of the fungus through the eggshell of the nematodes (**Bonants** *et al.* 1995). It is obvious that, *Paecilomyces lilacinus* is a cosmopolitan facultative pathogen of nematode juveniles and eggs (**Omaima** *et al.*, 2012). With regard to the bioagent compound, Sincocin, no reports are available in literature illustrating the role of Sincocin in controlling plant parasitic nematodes. Although the compound had a considerable effect on the nematodes, the doses used in this trial are considered so high. In addition, the non-trend of the concentrations tested may be due to that Sincocin contains the plant growth regulator, cytokinin which may

alter the susceptibility of the plant to nematode infection. Such alteration varied according to cytokin in concentration (Nyczepir and Wood, 1988). In relation to the bio- agent compound, Bio-Nema, Larvae of Meloidogyne spp. were readily infected with the endoparasite Bacillus penetrans by exposure to an aqueous suspension of spores from infected root-knot nematode females or by passage of larvae. Infection severely reduced motility of second-stage larvae or causes the death. Also, Shahnaz et al., (2008) reported that, application of Bacillus spp. significantly reduced hatching juveniles of *M. javanica* and mortality percentage of juveniles was significantly increased with increasing of exposure period.

 Table (2): Hatching percentage of *M. incognita* eggs as affected by different rates of the bio- agent compounds under laboratory conditions.

Exposure period	Treatment	Hatching %at different rates of application						
		1/4 Rate of application	1/2 Rate of application	Rate of application	2 Rate of application	4 Rate of application		
1 .4	Nembecidin	40.1	33.0	25.1	22.8	19.1		
1 uay	cadusafos	43.8	36.2	30.7	26.3	21.7		
	Bio-Nema	50.6	56.4	56.4	40.2	37.9		
7 days	Bio-Nematon	76.2	69.0	56.37	54.3	48.1		
	Sincocin	88.7	38.5	68.5	66.4	61.8		
	Nematoxin	43.3	34.8	28.4	24.4	20.9		

 Table (3): Mortality percentage of *M. incognita* J2 as affected by different rates of the bio- agent compounds under laboratory conditions.

Exposure period	T	Mortality% at different rates of application						
Exposure period	Treatment	1/4 Rate of application	1/2 Rate of application	Rate of application	2 Rate of application	4 Rate of application		
1 day	Nembecidin	25	64	71	81	89		
1 day	cadusafos	20	36	54	69	83		
	Bio-Nema	9	25	40	55	53		
7 days	Bio-Nematon	0	1	5	13	26		
	Sincocin	9	20	25	31	36		
	Nematoxin	61	68	80	85	96		

3.2. Greenhouse experiments.

3.2.1. Effect of application time and rate on the effectiveness of the tested bio-agent compounds against *M. incognita*.

Data given in Table (4) demonstrated the effect of the timing of application of the tested bio-agent compounds: Bio-Nema, Bio-Nematon, Sincocin, Nematoxin, Nembecidin and the chemical nematicide, cadusafos (G 10%) on the reproduction of nematode, *M. incognita*

www.esjpesticides.org.eg

infecting okra plants. Data illustrated that, all the tested compounds reduced number of galls, J2s in soil, number of eggs/ an eggmass, eggmasses and total number of eggs when applied before or after nematode inoculation. At the rate of application; treatment with Nematoxin when applied after nematode infection, caused the highest percentage of reduction for the numbers of galls, J2s in soil and total number of eggs (30.51, 38.20 and 30.10%, respectively), while treatment with Sincocin gave the lowest ones for the same parameters (1.81, 15.19 and 14.16%) respectively. On the other hand, treatment with Bio-Nematon when applied before nematode infection, gave the highest percentage of reduction for the number of galls, J2s in soil and total number of eggs (37.14, 55.83 and 48.40%) respectively, whereas treatment with Sincocin gave the lowest ones for the same parameters 32.66 and 24.18%), respectively. Generally, (29.99,application of the bio-agent compounds and the nematicide, cadusafos more effective when applied before nematode inoculate. With regard to the positive effect of the tested compounds comparing with their controls, on the growth of the infected okra plants with M. incognita, the data in Table (5) indicated that, treatment with Bio-Nema when applied before nematode infestation increased the length of shoot (7.9%), root (9.91%) and the fresh weight of shoot (22.48%) and root (16.44%) as well as total number of leaves (19.17%), in the while, increase comparing with the same comparative parameters with a percentage of 0.6, 1.29, 10.66, 0.34 and 0.0% respectively, when applied after nematode infestation. Also, data shown in Table (5) indicated that, there were a slightly differences among the bio-agent compounds on the growth parameters of okra plants within the same rate of application and in the same time. In contrast, the treatment with the nematicide, cadusafos, significantly increased the length of shoot (19.05%) and root (32.76%), fresh weight of shoot (59.94%) and root (57.53%) and number of leaves (37.5%) when applied before nematode infection, while the percentage of the same parameters increased about 13.69, 16.38, 49.86, 23.29 and 31.25%, respectively when applied after nematode infestation. However the present data are in agreement with Walia et al. (1991), who mentioned that, Paecilomyces lilacinus was applied either as a soil treatment 10 days before/after sowing as seed treatment or in different combinations to sandy soil planted with okra plants. Fungus aplication, in general, resulted in better top growth of okra, also, root galling was significantly reduced in all fungus treatments and proved better than carbofuran (1 kg a.i. /ha) in gall suppression. As mode of action of Paecilomyces lilacinus fungus, as observed by transmission electron microscopy, fungal hypha penetrated the M. javanica female cuticle directly (Khan et al. 2006). Mahgoob and El-Tayeb (2010) reported that, the nematicidal effect of the plant growth promoting bacteria (PGPB), Paenibacillus polymyxa UBF 15, Bacillus megaterium UBF 10 and B. circulans UBF 20 when applied two days before nematode-infestation was higher control compared with the application two days after infestation. Farahat et al. (1993) found that, the application of Sincocin after nematode inoculation significantly decreased numbers of the hatched juveniles and egg masses but without distinct trend. Double treatment of Sincocin gave the best results in reducing numbers of nematodes in soil. The addition of the compound one week after nematode inoculation resulted in a significant increase in shoot fresh and dry weights compared with the check, but without significant differences between concentrations. The failure of the high concentrations of Sincocin in reducing the numbersof nematodes may be due to cytokinin accumulation in plant roots may be justified by that cytokinin altered the resistant tomato cultivar, Y-91 to be susceptible to the root-knot nematode, M. igcognita (Dropkin et al., 1969).

Treatment			Nematode parameters (number / plant / pot)					
	Compounds		No. of	No. of J ₂ s	No. of	average eggs	Total No.	
Time		Rate of application	galls	In soil	eggmas	/an eggmas	of eggs	
	Bio-Nema		204.0d-g	7933.3b-e	163.5c-g	499.0c-h	81746.8	
	Bio-Nematon		215.3cde	8806.5b-e	176.3bcd	516.5b-f	91059.8	
	Sincocin		215.8cd	8807.5b-е	178.0bc	529.8a-d	94360.5	
	Nematoxin	1	191.8g-j	6417.8b-g	150.0gj	505.5c-g	75848.5	
	Nembecidin		200.8fgh	7389.5b-е	165.8c-f	488.5e-i	80952.8	
	cadusafos		66.00	1547.5ghi	42.00n	230.3p	9749.3	
	Bio-Nema		213.5c-f	8556.8b-e	173.8bcd	520.8а-е	90523.3	
	Bio-Nematon		219.0c	9243.8bcd	184.5b	531.8abc	98115.8	
er	Sincocin		240.0b	10345.8bc	184.5b	542.8ab	100183.3	
Aft	Nematoxin	1/2	213.5v-f	8045.0b-е	162.8d-g	501.3c-h	81718.8	
	Nembecidin		213.5c-f	8442.0b-е	171.8b-е	497.3d-h	85556.0	
	cadusafos		127.5m	3932.5d-r	70.0m	262.8p	18447.3	
	Bio-Nema		183.3jk	5522.5b-i	142.8h-k	405.0mno	58009.0	
	Bio-Nematon	2	205.0d-g	7263.0b-f	166.5c-f	515.3b-f	86323.3	
	Sincocin		203.5d-g	7705.0b-е	163.0d-j	471.5hij	77304.0	
	Nematoxin		182.0j-k	5478.0b-i	142.0ijk	382.50	54389.0	
	Nembecidin		185.8ijk	5708.0b-i	139.5jk	393.3no	54870.0	
	cadusafos		29.3.0b	570.0h-i	15.80	178.0q	2813.5	
	Bio-Nema		173.5k	4587.0c-i	138.3jk	410.3mno	56719.8	
-	Bio-Nematon		195.3g-i	1839.3a	139.5jk	421.51mn	58819.0	
	Sincocin		201.5e-h	6957.5b-f	161.5d-g	516.0b-f	83343.0	
	Nematoxin	1	197.5ghi	6603.0b-g	145.5hij	454.5jkl	66396.5	
	Nembecidin		159.0g-i	7633.8b-e	146.5hij	456.0ijk	67008.0	
	cadusafos		29.0p	799.0ghi	10.8o	178.5q	1939.0	
	Bio-Nema		198.3ghi	6479.5b-g	139.0jk	431.0klm	60026.5	
ъ	Bio-Nematon		203.3d-g	8138.3b-e	146.0hij	483.8f-i	70882.3	
for	Sincocin		223.3c	8729.0b-е	167.8c-f	516.5d-f	86733.8	
Be	Nematoxin	1/2	198.3ghi	8182.0b-е	145.5hij	494.0e-h	72079.5	
	Nembecidin		199.ghi	8221.3b-e	157.3eh	503.8c-h	79438.8	

Table (4): Effect of application time and the different rates of the tested bio - agent compounds on M.incognitainfecting okra plants.

In each column, means not followed by the same letter differ significantly from one another at the 0.05 level.

2

83.8n

154.81

168.0ijk

201.5e-h

188.8hij

186.3ijk

12.8u

276.0a

992.0ghi

3330.0e-i

4115.0d-i

6355.3b-h

4125.0d-i

4152.3d-i

10384.8b

237.5i

45.8n

122.01

130.5k

155.3f-i

130.5kl

139.0jk

198.6a

3.0o

243.0p

390.5no

479.0g-i

406.5mno

398.3mno

398.8mno

113.8r

552.6a

11167.0

47713.8

53095.5

74349.5

52024.0

55435.8

109921.8

343.0

cadusafos

Bio-Nema

Sincocin

Nematoxin

Nembecidin

Control inoculated untreated

cadusafos

Bio-Nematon

	Treatment	•	Plant parameters / plant					
T:	Commencedo	The rate of	Length (cm)		Fresh weight (gm)		No. of.	
TIME	Compounds	application	Shoot	Root	Shoot	Root	leaves	
	Bio-Nema		45.3efg	25.3de	85 fgh	3.4 fg	14.3 fgh	
	Bio-Nematon		44.5g-j	25.0def	8.5fgh	3.1g-h	14.3 fgh	
	Sincocin		42.5 l-o	23.0 ghi	6.7 klm	2.8 j	13.3 hij	
	Nematoxin	1	44.5 g-j	24.0 e-h	8.4 f-i	3.1 hij	13.3 hij	
	Nembecidin		44.3 g-k	24.5 d-g	7.8 g-k	3.1 hij	13.0 h-k	
	cadusafos		50.0 ab	30.8 a	11.1 bc	4.6 c	16.5 abc	
	Bio-Nema		43.3 j-n	23.5 f-i	7.7 g-l	3.2 ghi	13.8 ghi	
	Bio-Nematon		43.3j-n	23.0ghi	7.5h-m	3.1g-j	13.0h-k	
ore	Sincocin		42.3mno	22.8hi	4.6lm	2.9ij	11.8k	
Bef	Nematoxin	1/2	43.8h-l	23.3ghi	6.9j-m	3.0ij	11.8k	
	Nembecidin		43.3j-n	23.3ghi	6.9jm	2.9ij	13.3hij	
	cadusafos		48.8bv	29.5a	8.4fgh	4.1d	16.0b-e	
	Bio-Nema		46.3ef	25.3df	9.2def	3.7ef	15.0d-f	
	Bio-Nematon		45.0fgh	25.0def	9.0efg	3.6ef	15.3d-f	
	Sincocin		42.8l-o	23.3ghi	6.3m	3.0ij	13.3hij	
	Nematoxin	2	45.3efg	25.5cde	8.6e-h	3.4fgh	15.0d-f	
	Nembecidin		44.8ghi	25.0def	8.5fgh	3.5ef	14.8efg	
	cadusafos		50.5A	31.0a	11.2ab	5.0b	17.8a	
	Bio-Nema		42.25mno	23.5fi	7.68g-l	2.93ij	12.0jk	
	Bio-Nematon		42.25mno	23.25ghi	7.59h0m	2.92ij	12.25jk	
	Sincocin		42.51-o	23.25ghi	6.95j-m	2.92ij	12.25jk	
	Nematoxin	1	43.25j-n	23.5f-i	7.90g-k	2.97ij	12.5ijk	
	Nembecidin		43.0k-o	23.5f-i	7.89g-k	2.93ij	11.75k	
	cadusafos		47.75cd	27.0bc	10.4bcd	3.85de	15.75cde	
	Bio-Nema		42.25mno	23.0ghi	7.59h-m	2.92ij	12.0jk	
r	Bio-Nematon		42.25mno	22.25i	7.57h-m	2.92ij	12.0jk	
Afte	Sincocin		42.0no	23.0ghi	6.94j-m	2.92ij	12.25jk	
H	Nematoxin	1/2	43.25j-n	23.25ghi	7.76g-k	2.97ij	12.0jk	
	Nembecidin		42.751-o	23.5fi	7.44h-m	2.93ij	12.25jk	
	cadusafos		46.5de	25.75cd	9.87cde	3.55ef	15.5df	
	Bio-Nema		43.0k-o	24.0eh	8.13i-j	2.96ij	12.25jk	
	Bio-Nematon		43.0k-o	23.25ghi	7.65g-l	2.95ij	12.5ijk	
	Sincocin		42.51-o	23.25ghi	7.10i-m	2.93ij	12.0jk	
	Nematoxin	2	43.5j-n	24.0eh	8.28f-i	3.06hij	13.25hij	
	Nembecidin		43.5im	24.0eh	8.28f-i	3.02ij	13.0h-k	
cadusafos			48.0c	27.75b	10.25bc	4.05d	16.25bcd	
Control inoculated untreated			42.0o	32.2ghi	6.94j-m	2.92ij	12.0jk	
Control uninoculated untreated			50.75a	30.0a	12.46a	5.45a	17.25ab	

 Table (5): Effect of application time and the different rates of the tested bio- agent compounds on the growth of okra plants infected with *M. incognita*.
 on the different rates of the tested bio- agent compounds on the growth of okra plants infected with *M. incognita*.

In each column, means not followed by the same letter differ significantly from one another at the 0.05 level.

REFERENCES

- Abbott, W. S. (1925). A method of computing the effectiveness of an insecticide. Journal of Economic Entomology. 18: 265 – 267.
- Azam, M. F.; R. K Mehmood and A. Shamim (2001). Effect of plant extract of some members of Asteraceae on hatching and mortality of rootnematode. Meloidogyne knot incognita. Bionotes, 3: 9-10.
- Bonants P.J.M.; P.F.L. Fitters; E. Den Belder; C. Waal- wijik and J.W.D.M. Henfling (1995). A basic serine protease from Paecilomyces lilacinus with biological activity against Meloidogyne hapla eggs. Microbiology. 141: 775-784.
- Brand, D.; C. R. Soccol, A. Sabu, and S. Roussos. (2010). Production of fungal biological control agents through solid state fermentation: a case study on Paecilomyces lilacinus against rootknot nematodes. Micologia Aplicada International. 22: 31-48.
- hypersensitivity reaction of tomatoes resistant to Meloidogyne incognita reversal by citokinins. J. of Nematol. I: 55-61.
- Farahat 'A. A.; A. A. Osman; H. I. El-Nagar and H. H. Hendy (1993). Evaluation of Margosan and Sincocin as biocides of the reniform nematode infecting sunflower .Butt. Fac. of Agric. Univ. of Cairo. Vol. 44: 191-204.
- Goodey, J. B. (1957). Laboratory methods for work with plant and soil nematode. Tech. Bull. No. 2, Min. Agric., Fisheries & Food, London, 47 pp.
- Goodey, J. B. (1963). Laboratory methods for work with plant and soil nematodes. Tech. Bull. Minist. Agric. Fish. Fd. No. 2 (4th edition).
- Hussy, R. S. and K. R. Barker (1973). A comparison of methods of

collecting inocula of Meloidogyne spp., including a new technique. Pl. Dis. Reptr., 57 : 1925-1928.

- Ingham, R.; R. Dick and R. sattel (1999). Columbia root-knot nematode control in potato using corps. Oregon State univ. Extn - service publications, E M 8740, 8pp.
- Jatala P. (1986). Biological control of plant parasitic nematodes. Ann. Rev. Phytopathol. 24: 453-489.
- Jatala P., J. Franco; A. Gonzales and C.M. O'Hara (1985). Hatching stimulation and inhibition of Globodera pallida eggs by enzymatic and exopathic toxic compounds of some biocontrol fungi. J. Nematol.. 17: p. 501.
- A.; K.L. Williams and H.K.M. Nevalainen Khan, (2006). Control of plant-parasitic nematodes by Paecilomyce lilacinus and Monacrosporium lysipagum in pot trials. Biocontrol 51: 643-658.
- Khanna, A. S. and S. Kumar (2006). In vitro evaluation of neem-based nematicides against Meloidogyne incognita. Nematologia Mediterranea. 34: 49-54.
- Dropkin, V.II. Helgeson; J. P. and C.D. Upper (1969). The Kilama, P.; T. Dubois; D. Coyne; B. Niere; C. S.Gold and E. Adipala (2007). Antagonism of Paecilomyces spp. isolated from banana (Musa spp.) roots and rhizosphere against Radopholus similis. Nematropica, 37:215-225.
 - Linderman R.G. (1992). Vesicular-arbuscular mycorrhizae and soil microbial interactions. p. 45-70. In: "Mycorrhizae in Sustainable (G.J. Agriculture" Bethlenfalvay, R.G. Linderman, eds.). Amer. Soc. Agron. Special Publication 54. Amer. Soc. Agron., Madison, WI, 124 pp.
 - Mahgoob, A. E. and T. S. El-Tayeb (2010). Biological Control of Meloidogyne incognita on Tomato Using Plant Growth Promoting Bacteria. Egypt. J. Biol. Pest Control. 20: 95-103.

- Mankau R. and N. Prasad (1977).Infectivity of *Bacillus* penetrans in Plant-parasitic Nematodes. J. Nematol., 9: 40–45.
- Noling, J. W. and J. O Becker (1994). The challeng of research and extension to define and implement alternatives to methyl bromide. Journal of Nematology, 26: 573 – 786.
- Norton, D. C. (1978). Ecology of plant parasitic nematodes. Wiley, New York, 268 pp.
- Nyczepir, A. P. and B. W. Wood (1988). Peach leaf delayed by *Criconemella xenoplax*. J. of Nematol., 20: 585-589.
- Omaima, M. Hafez; Karima, H.E. Haggag; Wafaa. M.A. El-Nagdi and M.S. El-Shamma (2012). Effect of Potassium Fertilizer and Bio-Nematon on Performance of Williams Banana Cv. for Controlling Fungal Root Rot and Nematode Root-Knot Diseases. Journal of Applied Sciences Research, 8: 646-657.
- Prakob W.; V. Kanthasab; V. Supina; N. Chaimeungchern and T. Kidtayo (2007). Use of arbuscular mycorrhizal fungi, antagonistic fungus and rhizobacteria *P. aeruginosa* and *B.* subtillis in controlling tomato root-knot nematodes. J. Agric. 23: 403–406.
- Sasser, J. N. (1979). Economic important of *Meloidogyne* in tropical countries. From "Root-knot nematodes (*Meloidogyne* species) systemtics, biology and control". Ed. By F. Lamberti and C. E. Taylor. 477 pp.
- Shahnaz D.; T. Marium and M. J. Zaki (2008). Application of *Bacillus* species in control of *Meloidogyne javanica* (Treub) chitwood on cowpea and mash bean. Pakistan Journal of Botany. 40: 439-444.
- Sharma, G. C. (2000). Efficacy of neem based formulations against the root-knot nematode ' *Meloidogyne incognita* .Pesticide Research Journal. 12: 183-187.

- Siddiqui, I.A.; S. Ehetshamul-Haque and S. Shahid-Shaukat (2001). Use of rhizobacteria in the control of root rot and root knot disease complex of mungbean. J. Phytopathol. 149: 337–346.
- Walia, R. K.; R. K. Bansal and D. S. Bhatti (1991). Effect of *Paecilomyces lilacinus* application time and method in controlling *Meloidogyne javanica* on okra. Nematologia Mediterranea. 19: 247-249.

الملخص العربى

تقييم فاعلية بعض المركبات الحيوية مقارنة بمبيد الكادوسافوس ضد نيماتودا تعقد الجذور (ميلودوجيني إنكوجنيتا)

أحمد عيد عبد المجيد محجوب٬، سيد عبد اللطيف محمد دحروج٬ ، أحمد عبد المحسن أحمد دعبس٬ ،عايدة محمد الخولي٬

١ قسم وقاية النبات – كلية الزراعة – جامعة عين شمس– – شبرا الخيمة

٢– قسم بحوث المبيدات الفطرية و البكتيرية و النيماتودية - المعمل المركزى للمبيدات – مركز البحوث الزراعية – الجيزة

أجريت هذه الدراسة بهدف إختبار فاعلية بعض المركبات الحيوية مثل بيونيما ، بيونيماتون ، نيماتوكسين ، نيمبسيدين وسينكوسين ، مقارنة بالميبد النيماتودي كادوسافوس ضد نيماتودا تعقد الجذور ميلودوجيني إنكوجنيتا وذلك للحد من تأثيرها الضار على النبات بالإضافة إلى تحسن نمو النبات والحفاظ على البيئة ، وقد تم إجراء هذه الدراسة على مرحلتين كما يلي:

اختبارات معملية: أجريت تجربتين كالآتى:

أ- إختبار فاعلية هذه المركبات على فقس بيض نيماتودا تعقد الجذور ميلودوجينى إنكوجنيتا ، وفى هذه التجربة تم إستخدام خمس معدلات وهى: معدل التطبيق ، ربع معدل التطبيق ، نصف معدل التطبيق ، وضعف معدل التطبيق ، و ضعفى معدل التطبيق ، وذلك عن طريق إضافة كل معدل إلى معلق يحتوى على ١٠٠٠ بيضة نيماتودا ثم فحص البيض بعد يوم واحد لكل من مركب نيمبسيدين و مبيد كادوسافوس ، وبعد لاأيام لكل من المركبات بيونيما ، بيونيماتون ، وسينكوسين و النيماتوكسين وذلك لتقدير نسبة فقس البيض ، وأظهرت النتائج مايلى:

۱- بعد يوم واحد من المعاملة كانت أقل نسبة فقس هي ۱۹٫۱ و ۲۱٫۷ % عند ضعفي المعدل المقترح وأعلى نسبة فقس كانت ٤٠٫١ و ٤٣٫٨ عند ربع معدل التطبيق من النيمبسيدين و كادوسافوس على التوالي.

٢- بعد ٧ أيام من المعاملة كانت أقل نسبة ٢٠,٩ و ٣٧,٩ عند ضعفى معدل التطبيق لكل من النيماتوكسين و البيونيما على التوالى بينما كان أعلى معدل للفقس ٨٨,٧ و ٨٣,٥ % عند ربع و نصف معدل التطبيق معدل التطبيق لمركب السينكوسين على التوالى و ٧٦,٢ % عند ربع معدل التطبيق لمركب البيونيما.

ب- إختبار فاعلية هذه المركبات على موت الطور اليرقى الثاني لنيماتودا تعقد الجذور ميلودوجيني إنكوجنيتا.

في هذه التجربة تم إختبار نفس المعدلات السابقة لنفس المركبات وفترات الفحص ولكن تم إضافة هذه المعدلات إلى معلق يحتوى على عدد ٣٠٠ برقة نيماتودا وتم حساب نسبة الموت وكانت النتائج كالآتي:

١- بعد يوم من المعاملة كانت أعلى نسبة موت هي ٨٣,٨٩ % عند ضعفي معدل التطبيق وأقل نسبة موت هي ٢٠,٢٥ عند ربع معدل التطبيق لكل من مركب النيمبسيدين و مبيد كادوسافوس على التوالي.

۲ ـ بعد ۷ أيام من المعاملة كانت نسبة الموت ٥٣ و ٩٦% عند ضعف معدل التطبيق لمركبي بيونيما و نيماتوكسين على التوالي بينما أقل نسبة موت كانت ٥,٥ و ٩ % عند ربع معدل التطبيق لكل من بيونيما و بيونيماتون على التوالي.

٢ - إختبارات نصف حقلية تحت ظروف الصوبه.

في هذه الإختبارات تم تقييم إختبار توقيت إضافة المركبات على فاعليتها ضد نيماتودا تعقد الجذور ميلودوجيني إنكوجنيتا.

في هذه التجربة تم إختبار ٣ معدلات من كل مركب (نصف معدل التطبيق – معدل التطبيق – ضعف معدل التطبيق) وتم إضافتها مرة قبل عدوى النباتات بمدة أسبوع و مرة أخرى بعد عدوى النباتات بمدة أسبوع على مجموعتين مختلفتين من النباتات وكانت النتائج كالآتي:

١- عند إضافة المركبات بعد عدوى النباتات كانت أعلى نسبة خفض فى تعداد العقد الجذرية و الطور اليرقى الثانى فى التربة و العدد الكلى للبيض هى (
 ٣٠,٥١ و ٣٨,٢٠ و ٣٠,١٠) على التوالى عند معدل التطبيق لمركب نيماتوكسين ، بينما كانت أقل نسبة خفض (٢١,٨١ و ١٥,١٩ و ١٤,١٦ %)
 لنفس مظاهر النشاط السابقه على التوالى عند معدل التطبيق لمركب السنكوسين.

٢- عند إضافة المركب قبل عدوى النباتات بالنيماتودا كانت أعلى نسبة خفض لتعداد العقد الجذرية ، الطور البرقى الثاني في التربه والعدد الكلى للبيض هي (٣٢,٦٤ و ٣٢,٦٦ و ٣٢,٦٦ و ٣٢,٦٦ و ٣٢,٦٦ و ٣٢,٦٦ و ٣٢,٦٦

٣- مبيد كادوسافوس أعطى أعلى نسبة خفض على الإطلاق في تعداد العقد الجذرية والطور البرقي الثاني في التربة والعدد الكلي للبيض (٨٩,٤٩ و ٩٢,٣٠ و ٩٨,٢٤ %) على التوالي عند إستخدام المعدل الموصى به قبل عدوي النباتات بالنيماتودا.

٤ ـ أظهرت إضافة المركبات قبل العدوى بالنيماتودا زيادة ملحوظة في طول المجموع الجذري والخضري وكذلك وزنها مقارنة بالنباتات المصابة والغير معاملة بالمركبات.